Synthesis of Hyperbranched Polytetritol by Ring-Opening Multibranching Polymerizations of 2,3-Anhydroerythritol and 2,3-Anhydro-DL-threitol

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Received September 13, 2004; Revised Manuscript Received November 25, 2004

ABSTRACT: 2,3-Anhydroerythritol ($1\mathbf{a}$) and 2,3-anhydro-DL-threitol ($1\mathbf{b}$) were polymerized using boron trifluoride diethyl etherate (BF₃-OEt₂) as a cationic initiator. The polymerizations of $1\mathbf{a}$ and $1\mathbf{b}$ proceeded through a ring-opening reaction with a proton-transfer reaction to produce hyperbranched carbohydrate polymers ($2\mathbf{a}$ and $2\mathbf{b}$) consisting of DL-threitol and erythritol units, respectively. The degrees of branching (DBs) estimated by the ¹³C NMR spectra of $2\mathbf{a}$ and $2\mathbf{b}$ were 0.47 and 0.45, respectively. The weight-average molecular weight ($M_{\text{w,SLS}}$) values ($2.67 \times 10^5 - 3.20 \times 10^6$) estimated using static light scattering (SLS) of the resulting hyperbranched carbohydrate polymers were significantly higher than the weight-average molecular weight ($M_{\text{w,SEC}}$) values ($1.04 \times 10^3 - 2.77 \times 10^3$) estimated using size exclusion chromatography (SEC). The viscosities of $2\mathbf{a}$ and $2\mathbf{b}$ in aqueous sodium nitrate (NaNO₃) solution were very low, and the intrinsic viscosities ($[\eta]$) of $2\mathbf{a}$ and $2\mathbf{b}$ were in the range from 0.0190 to 0.0250 dL g⁻¹. The three-dimensional properties characterized by the SLS and viscosity measurements indicated that $2\mathbf{a}$ and $2\mathbf{b}$ should be spherical molecules.

Introduction

Ring-opening multibranching polymerization of cyclic monomers is a versatile method for synthesizing hyperbranched macromolecular architectures. Since Suzuki et al. first reported the ring-opening multibranching polymerization using a cyclic carbamate, 1a,b there have been were various studies using cyclic monomers, such as oxetanes, ^{2a-d} tetrahydrofurans, ³ lactones, ^{4a,b} and oxiranes. ^{5a-e} In particular, glycidol is the simplest, commercially available, and cheap oxirane, so that many efforts to prepare hyperbranched polyglycerol have been carried out; for example, Penczek and Dworak et al. reported the synthesis of a highly branched polymer by the cationic polymerization of glycidol. 5a,b In addition, Frey et al. reported the anionic ring-opening multibranching polymerization of glycidol and used the resulting hyperbranched polyglycerol as the molecular nanocapsules for hydrophilic guests. 5c-e Thus, for the ring-opening multibranching polymerization using cyclic monomers, the design and synthesis of hyperbranched polymers with novel structures together with their application is of significant interest.

Previously, we proposed a hyperbranched carbohydrate polymer as a novel class of polysaccharides, which was based on the synthetic strategy of the ring-opening multibranching polymerization of anhydro sugars as cyclic monomers, such as 1,2:5,6-dianhydro-D-mannitol,⁶

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1,6-anhydro- β -D-mannopyranose, ^{7a,b} 1,6-anhydro- β -Dglucopyranose, 7b and 1,6-anhydro-β-D-galactopyranose. 7b The resulting hyperbranched carbohydrate polymers exhibited characteristic properties due to their spherical and compact structure, e.g., a high water solubility and extremely low solution viscosity, which significantly differed from those of linear carbohydrate polymers. To expand the family of hyperbranched carbohydrate polymers, it is important to elucidate applicable anhydro sugar monomers for the ring-opening multibranching polymerization. Recently, we reported the synthesis of hyperbranched polytetritol by the ring-opening multibranching polymerization of 1,4-anhydroerythritol (3a) and 1,4-anhydro-L-threitol (3b).3 Although these monomers were successfully polymerized using superacids, such as trifluoromethanesulfonic acid and fluorosulfonic acid, as the cationic initiator, the resulting hyperbranched polytetritol contained the 1,4-anhydrotetritol unit as an unexpected unit. Our continuous interest is to expand the family of hyperbranched carbohydrate polymers along with their characteristic properties due to their spherical and compact structure, so that it is important to synthesize and characterize hyperbranched polytetritol without any anhydrotetritol unit. Thus, we now report the synthesis of a hyperbranched carbohydrate polymer (3a, 3b) essentially consisting of tetritol units by the ring-opening multibranching polymerization using 2,3-anhydroerythritol (1a) and 2,3-anhydro-DL-threitol (1b), which are the structural isomers of 1,4anhydrotetritol, as shown in Scheme 1. The structures of **2a** and **2b** are characterized using the results based on the NMR, static light scattering, and solution viscosity measurements by the comparison with a linear polymer of $(2\rightarrow 3)$ -erythritol (4) along with hyperbranched polytetritol prepared from 3a and 3b. In

Scheme 1

HO OH
$$\frac{BF_3 \cdot OEt_2}{1a}$$
 $\frac{BF_3 \cdot OEt_2}{1b}$ $\frac{BF_3 \cdot OEt_2$

addition, the polymerization mechanisms of 1a and 1b are discussed on the basis of the structures and the degrees of branching (DBs) of the resulting hyperbranched poly-DL-threitol (2a) and hyperbranched polyerythritol (2b).

3b

Experimental Section

3a

Measurements. The ¹H (400 MHz) and ¹³C NMR spectra (100 MHz) were recorded using a JEOL JNM-A400II instrument. The quantitative ¹³C NMR spectra were obtained using a 20% (w/v) sample in deuterium oxide (D₂O) at 25 °C, 45° pulse angle, inverse gated decoupling with a 7.0 s delay, 6000 scans, and acetone as the internal reference. Size exclusion chromatography (SEC) for the water-soluble polymer was performed in aqueous sodium nitrate (NaNO₃) solution (0.2 mol L⁻¹) at 40 °C (1.0 mL min⁻¹) using a TOSOH HPLC system (HLC-8020) equipped with two TSK_{gel} GMPW_{XL} columns (pore size, 12.5–100 nm; bead size, 13 μ m; exclusion limit, 5×10^7) and a refractive index detector. The weightaverage molecular weights $(M_{\rm w,SEC})$ and the molecular weight distributions (M_w/M_n) of the polymer samples were calculated on the basis of poly(ethylene glycol) calibrations. For the organic solvent-soluble polymer, the SEC measurement was performed at 40 °C in chloroform (0.8 mL min⁻¹) using a Jasco GPC-900 system equipped with a TOSOH TSK_{gel} GMH_{HR}-M column (linear, $7.8 \text{ mm} \times 300 \text{ mm}$; pore size, 16 nm; bead size, 5 μ m; exclusion limit, 4 \times 106), a Shodex KF-804L column (linear, 8 mm \times 300 mm; pore size, 20 nm; bead size, 7 μ m; exclusion limit, 4×10^5), and a refractive index detector. The $M_{\rm w,SEC}$ and the $M_{\rm w}/M_{\rm n}$ were calculated on the basis of a polystyrene calibration. The preparative SEC was performed in chloroform (3.8 mL min⁻¹) at 23 °C using a JAI LC-908 equipped with JAI JAIGEL-2H polystyrene column (pore size, 40-50 Å; bead size, 15 μ m; exclusion limit, 5×10^3), a JAIGEL-2.5H polystyrene column (pore size, 60-70 Å; bead size, 16.5 μ m; exclusion limit, 2 \times 10⁴), and JAI UV-310 and JAI RI-5HC detectors. A static light scattering (SLS) measurement was performed in an aqueous $NaNO_3$ solution (0.2 mol L^{-1}) or toluene at 40 °C using an Otsuka Electronics CALLS-1000 light scattering spectrometer ($\lambda = 632.8$ nm). The refractive index increment (dn/dc) was measured in aqueous NaNO3

solution (0.2 mol L⁻¹) or toluene using an Otsuka Electronics DRM-1021 double-beam differential refractometer. The intrinsic viscosity ([\eta]) was determined in aqueous NaNO3 solution (0.2 mol L⁻¹) at 40 °C using a Canon-Fenske viscometer after the sample solution was filtered.

Materials. cis-2-Butene-1,4-diol (Tokyo Kasei Co., >94%), 2-butyne-1,4-diol (Kanto Kagaku Co., 97%), N,N-dimethylformamide (DMF) (Junsei Chem. Co., 99.0%), anhydrous magnesium sulfate (Junsei Chem. Co., 99.5%), methanol (Junsei Chem. Co., 99.6%), acetone (Junsei Chem. Co., 99.0%), chloroform (Junsei Chem. Co., 98.0%), allyl bromide (Kanto Kagaku Co., 99.0%), ethyl alcohol (Japan Alcohol Trading Co., 99.0%), and silver oxide (Ag₂O) (Aldrich, 99.0%) were used without further purifications. Boron trifluoride diethyl etherate $(BF_3 \cdot OEt_2)$ (Kanto Kagaku Co., 95.0%) was purified by distillation under reduced pressure. The dialysis membranes (Spectra/Por6 regenerated cellulose, MWCO 1000, and Spectra/ Por cellulose ester MWCO 500) were purchased from Spectrum Laboratories, Inc. 2,3-Anhydroerythritol (1a) and 2,3-anhydro-DL-threitol $(\mathbf{1b})$ were synthesized from $\mathit{cis} ext{-}2 ext{-}\mathrm{butene} ext{-}1 ext{,}4 ext{-}\mathrm{diol}$ and trans-2-butene-1,4-diol,8 respectively, according to the methods from a previous report. 9a,b

Synthesis of 2,3-Anhydro-1,4-di-O-allylerythritol (5). To a solution of 2,3-anhydroerythritol (3.33 g, 31.7 mmol) in DMF (50 mL) were added silver oxide (20.0 g, 86.3 mmol) and allyl bromide (10.0 mL, 116 mmol) at 40 °C. The reaction mixture was stirred at 40 °C for 4 h. The suspension was cooled, and the salts were filtered off and washed with acetone. The filtrates were evaporated under reduced pressure. The residue was purified by column chromatography on silica gel with hexane/ethyl acetate (10:1 v/v) and distilled under reduced pressure to give a colorless liquid 5 (2.2 g) in a 37.4% yield; bp 141 °C/33 mmHg; R_f 0.29. ¹H NMR (CDCl₃): $\delta = 5.91 - 5.81 \, (\text{m, allyl, -CH=, 2H}), \, 5.24 \, (\text{ddd, }^3J_{\text{trans}} = 17.3 \, \text{Hz}, \, J_{\text{gem}} = 3.2 \, \text{Hz, }^4J = 1.7 \, \text{Hz}, \, \text{allyl trans} = \text{CH}_2, \, 2\text{H}), \, 5.16$ (ddd, ${}^{3}J_{cis} = 10.5 \text{ Hz}$, $J_{gem} = 3.2 \text{ Hz}$, ${}^{4}J = 1.2 \text{ Hz}$, allyl, cis = CH_2 , 2H), 4.06-3.93 (m, allyl, $-CH_2$ -, 4H), 3.64 (dd, J_{gem} = 17.3 Hz, $J_{\text{vic}} = 3.4$ Hz, CH₂, H-1 and H-4, 2H), 3.49-3.46(m, CH₂, H-1 and H-4, 2H), 3.20-3.16 ppm (m, epoxy, CH, H-2 and H-3, 2H). ¹³C NMR (CDCl₃): $\delta = 134.29$ (allyl, -CH=), 117.43 (allyl, =CH₂), 72.14 (allyl, -CH₂-), 68.03 (CH₂), 54.38 ppm (CH). Anal. Calcd for C₁₀H₁₆O₃ (184.2): C, 65.19; H, 8.75. Found: C, 64.99; H, 8.78. FI-MS: *m/z* (relative intensity): 184 $(M^+, 100), 185 (MH^+, 79.4).$

Polymerization of 5. All procedures were performed under an argon atmosphere. The monomer **5** (1.00 g, 5.43 mmol) was placed in an ampule and dried under reduced pressure for 18 h. After adding BF₃·OEt₂ (34.4 μL, 0.271 mmol), the ampule was sealed and cooled at 0 $^{\circ}\text{C}$ for 240 h. The polymerization mixture was quenched by adding water (5 mL) and then neutralized with aqueous ammonium solution. The mixture was extracted three times with chloroform, dried with anhydrous magnesium sulfate, filtered, and evaporated. The residue was dissolved in chloroform (1 mL). A part of the solution (0.2 mL) was applied to the preparative SEC, and then the eluents with the retention times of 30-45 min were fractionated. This procedure was repeated five times. The combined fractions were evaporated and then dried in vacuo to give $(2\rightarrow 3)$ -1,4-di-O-allyl-DL-threitol (6) (0.720 g) in a 72.0% yield. The $M_{\rm w,SEC}$ and $M_{\rm w}/M_{\rm n}$ values were 1800 and 1.40, respectively. ¹H NMR (CDCl₃): $\delta = 5.87-5.60$ (m, allyl, -CH=, 2H), 5.25 (d, $J_{\text{vis}} = 17.1 \text{ Hz}$, allyl, trans =CH₂, 2H), 5.25–4.84 (m, allyl, =CH₂, 2H), 3.94-3.75 (m, allyl, -CH₂-, 4H), 3.73-3.01 (m, CH and CH₂, 6H). ¹³C NMR (CDCl₃): $\delta = 134.68$ (allyl, -CH=), 116.11 (allyl, =CH₂), 77.48 (C2 and C3), 71.78 (allyl, -CH₂-), 69.29 ppm (C1 and C4).

Isomerization of 6. To a solution of **6** (0.430 g) in toluene (70 mL), ethanol (160 mL), and water (2 mL) was added tris-(triphenylphosphine)chlororhodium (RhCl(PPh₃)₃) (0.364 g, 0.393 mmol) and 1,4-diazabicyclo[2.2.2]octane (DABCO) (0.130 g, 1.16 mmol). After the reaction mixture was refluxed for 12 h, it was cooled to room temperature, and the solvent was evaporated. The residue was purified by the preparative SEC as well as 6. The combined fractions were evaporated and then

Table 1. Polymerization of 2,3-Anhydrotetritol (1a and 1b) Using Borontrifluoride Diether Etherate

run	monomer	[M]/[I]	temp/°C	time/h	yield/%	$M_{ m w,SEC} (M_{ m w}/M_{ m n})^a$	$M_{ m w,SLS}^b imes 10^{-5}$	$[\eta]^c imes 10^{-2} / \mathrm{dL}~\mathrm{g}^{-1}$	DB^d
1	1a	100	80	120	44.8	1040 (2.32)	2.67	1.90	0.47
2	1a	100	100	60	41.2	1310 (1.87)	3.38	1.99	0.47
3	1a	100	100	100	57.3	1770 (1.99)	16.1	2.30	0.47
4	1a	100	100	120	60.8	2170 (2.98)	31.2	2.48	0.47
5	1a	200	100	120	51.8	2770(2.45)	32.0	2.50	0.47
6	1b	100	100	120	61.4	2400 (2.55)	30.8	2.42	0.45

 a Measured in an aqueous NaNO $_3$ solution (0.2 mol L $^{-1}$) using poly(ethylene glycol) as standards. b Measured in an aqueous NaNO $_3$ solution (0.2 mol L $^{-1}$). c Obtained from Canon-Fenske viscometer in an aqueous NaNO $_3$ solution (0.2 mol L $^{-1}$). d Determined from 13 C NMR spectra.

dried in vacuo to give (2 \rightarrow 3)-1,4-di-O-2-propenyl-DL-threitol (7) (0.380 g) in a 88.4% yield.

Hydrolysis of 7. To a solution of 7 (380 mg) in acetone (180 mL) was added a 0.5 mol L⁻¹ aqueous HCl solution (6 mL), and the mixture was heated under reflux for 15 min. The mixture was evaporated, and the residue was purified using a dialysis membrane (MWCO 500) in water. After dialysis, the residue was freeze-dried to give a white solid of (2 \rightarrow 3)-DL-threitol (4) (85.0 mg) in a 39.6% yield. The $M_{\rm w,SEC}$ and $M_{\rm w}/M_{\rm n}$ values were 890 and 1.22, respectively, and the $M_{\rm w,SLS}$ value was $1.08 \times 10^3 \, ({\rm d}n/{\rm d}c = -5.91 \times 10^{-1})$. ¹H NMR (D₂O) δ : 4.14-3.37 ppm (broad). ¹³C NMR (D₂O) δ : 81.81 (C2 for the terminal unit), 79.72 (C2 and C3 for the repeating unit), 76.34 (CH₂), 71.93 (C3 for the terminal unit), 67.43 (CH₂), 62.84 (C4 for the terminal unit), 60.99 ppm (C1 and C4 for the repeating unit and C1 for the terminal unit).

Scheme 2

Ag₂O

DMF

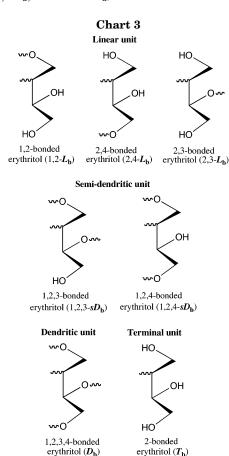
$$A_{g_2O}$$
 A_{g_2O}
 $A_{g_$

Typical Procedure for Polymerization of 2,3-Anhydroerythritol (1a). All procedures were performed under an argon atmosphere. Monomer 1a (1.00 g, 9.61 mmol) was placed in an ampule and dried under reduced pressure for 18 h. The ampule was heated at 100 °C, and then BF₃·OEt₂ (12.2 μL, 9.61×10^{-2} mmol) was added. After 120 h, the polymerization mixture was quenched by adding water (5 mL), then neutralized with an aqueous ammonia solution, and evaporated. The residue was dissolved in methanol (1 mL) and then poured into a large volume of acetone. The precipitated polymer was filtered off, and the filtrate was purified using a dialysis membrane (MWCO 500) in water. After dialysis, the residue was freeze-dried to give the polymer of 2a (0.608 g) in a 60.8% yield. The $M_{\rm w,SEC}$ and $M_{\rm w}/M_{\rm n}$ values were 2.17×10^3 and 2.98, respectively, and the $M_{
m w,SLS}$ value was $3.12 imes 10^6 \, ({
m d}n/{
m d}c = 1.40$ \times 10⁻¹). ¹H NMR (D₂O) δ : 4.41–4.33 (m), 4.06–3.89 (m), 3.85–

3.69 (m, H-3 for 2-bonded DL-threitol (T_a) as the terminal unit), 3.68–3.52 (m), 3.50–3.37 ppm (m, H-2 for T_a). 13 C NMR (D₂O) δ : 85.61 (CH), 81.76 (C2 for T_a), 81.20–80.45 (CH₂), 80.24 (C2 and C3 for 2,3-bonded-DL-threitol (2,3- L_a) as the linear unit), 75.64 (CH), 74.32 (CH), 73.60 (CH₂), 72.98–72.81 (CH), 72.34 (C3 for T_a), 71.18 (CH₂), 71.15–69.55 (CH), 69.42–67.61 (CH₂), 63.40 (C4 for T_a), 62.00–61.13 (CH₂), and 60.44 ppm (C1 for T_a and C1 and C4 for 2,3- L_a).

Procedure for Polymerization of 2,3-Anhydro-DL-threitol (1b). All procedures were performed under an argon atmosphere. Monomer 1b (1.00 g, 9.61 mmol) was placed in an ampule and dried under reduced pressure for 18 h. The ampule was heated at 100 °C, and then BF₃·OEt₂ (12.2 μ L, 9.61 \times 10⁻² mmol) was added. After 120 h, the polymerization mixture was quenched by adding water (5 mL), neutralized

with an aqueous ammonium solution, and then evaporated. The residue was dissolved in methanol (1 mL) and then poured into a large amount of acetone. The precipitated polymer was filtered off, and the filtrate was purified using a dialysis membrane (MWCO 500) in water. After dialysis, the residue was freeze-dried to give the polymer of 2b (0.614 g) in a 61.4% yield. The $M_{\rm w.SEC}$ and $M_{\rm w}/M_{\rm n}$ values were 2.40×10^3 and 2.55, respectively, and the $M_{
m w,SLS}$ value was $3.08 imes 10^6 \, ({
m d}n/{
m d}c = 1.04$ \times 10⁻¹). ¹H NMR (D₂O) δ : 4.46–4.34 (m), 4.22–4.20 (m, H2 and H3 for 2,3-bonded erythritol (2,3-Lb) as the linear unit, 2H), 4.09-3.65 (m), 3.52-3.31 ppm (m, H2 for 2-bonded erythritol (T_b) as the terminal unit). ¹³C NMR (D_2O) δ : 83.01 (C2 for T_b), 81.80 (C2 and C3 for D_b), 81.49 (C2 and C3 for $2,3-L_b$, 76.88 (CH), 74.94 (CH), 74.24 (CH₂), 74.01 (CH), 73.38(CH), 73.00 (CH₂), 72.69 (C3 for T_b), 71.98 (CH), 71.64 (CH), 71.27 (C1 and C4 for **D**_b), 70.89 (CH), 70.44-69.29 (CH₂), 65.85 (CH₂), 64.77 (C4 for T_b), 64.31 (CH₂), and 60.11 ppm (C1 and C4 for 2,3- L_b , and C1 for T_b).



Degree of Branching (DB). The DBs of 2a and 2b prepared from the AB3 monomer were determined from the ratio of the terminal units using Frey's equation.3,6,10 In the ¹³C NMR spectrum of **2a**, the signal at 72.34 ppm was assigned to the C3 methyne carbon due to the 2-bonded DL-threitol (T_a) as the terminal unit. The ratio of T_a , $T_a/(D_a + sD_a + L_a +$ $T_{\rm a}$), was estimated from the integrated area of the methyne carbon peaks as $[2 \times (\text{the integrated area of C3 methyne peak}]$ for T_a)/(the integrated area of total methyne peaks)]. The DB of polymer 2a was calculated as shown in eq 1.

$$DB = \frac{3}{2} \left| \frac{T_a}{D_a + sD_a + L_a + T_a} \right|$$
 (1)

This calculation method was applied to 2b. From the ratio of the 2-bonded erythritol ($T_{\rm b}$) as the terminal unit, which was estimated from the signal at 72.69 ppm in the $^{13}\mathrm{C}$ NMR spectrum, the DB of 2b was calculated.

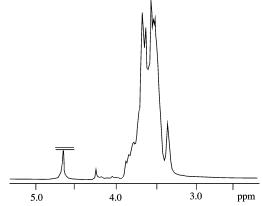


Figure 1. ¹H NMR spectrum of 2a (run 4).

Results and Discussion

Polymerization. The bulk polymerization of 2,3anhydroerythritol (1a) and 2,3-anhydro-DL-threitol (1b) was carried out using boron trifluoride diethyl etherate (BF₃·OEt₂) as the initiator at 80 or 100 °C. After the initiator was added to the monomer, the viscosity of the polymerization system immediately increased. The polymeric products were isolated by reprecipitation using methanol and acetone. The resulting polymers were white solids, which were soluble in water, sparingly soluble in methanol, and insoluble in other organic solvents. Table 1 summarizes the polymerization results. For the polymerization of 1a, the weight-average molecular weight values $(M_{
m w,SLS})$ measured by the static light scattering (SLS) and the yield of the resulting polymer increased with the rising the polymerization temperature; i.e., when the [1a]/[BF₃·OEt₂] molar ratio of 100 was used for 120 h, the $M_{\rm w.SLS}$ value was 2.67 \times 10^5 in 44.8% yield at 80 °C (run 1) and 1.61×10^6 in 57.3% yield at 100 °C (run 3). The $M_{\rm w.SLS}$ value and yield also increased with the extension of the polymerization time (runs 2, 3, and 4). For the polymerization of **1b** at 100 °C for 120 h, the $M_{\rm w.SLS}$ values were 3.08×10^6 in 61.4% yield in the $[1b]/[BF_3 \cdot OEt_2]$ molar ratio of 100. The $M_{\rm w,SLS}$ values (2.67 imes $10^5-3.20$ imes 10^6) of the resulting polymers were higher than those $(1.00 \times 10^4 5.10 \times 10^{5}$) of the hyperbranched polytetritols prepared by the ring-opening multibranching polymerizations of 1,4-anhydroerythritol (3a) and 1,4-anhydro-L-threitol $(3b).^3$

Polymer Structure and Degree of Branching. In the ¹H NMR spectrum of the polymer obtained from **1a**, the characteristic absorption at 3.25–3.35 ppm due to the epoxy proton of **1a** disappeared, as shown in Figure 1, indicating that the epoxy group in **1a** was consumed. Thus, the polymer obtained from 1a should consist of 14 kinds of DL-threitol repeating units (Chart 2). To investigate the polymer structure, the ${}^{13}\mathrm{C}$ NMR spectrum of the polymer was compared with that of a linear carbohydrate polymer of (2→3)-DL-threitol (4) as shown in Figure 2. For the ¹³C NMR spectrum of 4, the major signals marked by the open circles at 79.72 and 60.99 ppm were assigned to the C2,3 and C1,4 carbons for the 2,3-bonded DL-threitol repeating unit, respectively. In the comparison with the chemical shifts of L-threitol as the model compound,³ the small signals marked by the closed circles at 81.81, 71.93, 62.84, and 60.99 ppm were assigned to the C2, C3, C4, and C1 carbons for the 2-bonded DL-threitol unit as the polymer terminals, respectively. On the other hand, the ¹³C NMR spectrum 3of the polymer obtained from **1a** consisted of a number of split and broad signals. Some signals correspond to those of 4; e.g., the major signals marked by the open squares at 81.76, 72.34, 63.44, and 60.44 ppm agreed with the chemical shifts of the signal for the 2-bonded DL-threitol terminal unit. The small signals marked by the closed squares at 80.24 and 60.44 ppm agreed with the chemical shifts of the signal for the 2,3-bonded DLthreitol repeating unit of 4. In addition, by comparison with the hyperbranched poly-L-threitol prepared from **3b**, the remaining signals could be assigned to the 1,2bonded DL-threitol $(1,2-L_a)$ as the linear unit, 1,2,3- and 1,2,4-bonded-DL-threitol $(1,2,3-sD_a)$ and $(1,2,4-sD_a)$ as the semi-dendritic unit, and 1,2,3,4-bonded-DL-threitol $(1,2,3,4-D_a)$ as the dendritic unit. Thus, the polymer obtained from 1a was a hyperbranched poly-DL-threitol (2a) containing numerous 2-bonded DL-threitols (T_a) as the terminal units.

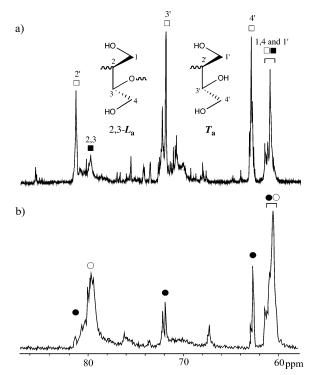


Figure 2. ^{13}C NMR spectra of (a) **2a** (run 4) and (b) (2 \rightarrow 3)-DL-threitol (3).

For the polymer obtained from 1b, the ¹H NMR and $^{13}\mathrm{C}$ NMR spectra are shown in Figure 3. In the $^{1}\mathrm{H}$ NMR spectrum, the signal was broad and the characteristic absorption at 3.25-3.35 ppm due to the epoxy proton of 1b disappeared, so that the polymer obtained from 1b should consist of 7 kinds of erythritol repeating units as shown in Chart 3. The ¹³C NMR spectrum consisted of a number of split and broad signals. In the comparison with the chemical shifts of erythritol as the model compound, the major signals marked by the open triangles at 83.01, 72.69, 64.77, and 60.11 ppm were assigned to the carbon for the 2-bonded erythritol ($T_{
m h}$) as the terminal unit.³ The shape of the ¹³C NMR spectrum of the polymer obtained from 1b was similar to that of 2a, so that the signals marked by the closed triangles at 81.49 and 60.11 ppm should be assigned to the carbon for the 2,3-bonded erythritol $(2,3-L_b)$ as the linear unit. By comparison with the hyperbranched polyerythritol prepared from **3b**, ³ the rest of the signals could be assigned to 1,2-bonded erythritol $(1,2-L_b)$ as

the linear unit, 1,2,3- and 1,2,4-bonded erythritol (1,2,3- and 1,2,4- $\mathbf{s}D_{\mathbf{b}}$), and 1,2,3,4-bonded erythritol ($D_{\mathbf{b}}$) as the dendritic unit. For example, the signals marked by the open diamonds at 81.80 and 71.27 ppm were assigned to the carbon for $D_{\mathbf{b}}$. These results indicated that the polymer obtained from $\mathbf{1b}$ should be a hyperbranched polyerythritol ($\mathbf{2b}$) having numerous erythritol terminal units.

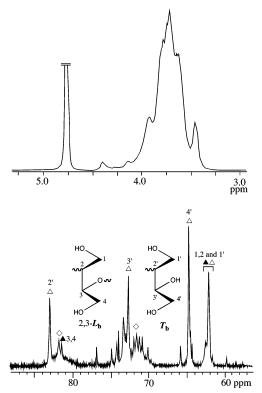


Figure 3. (a) ¹H NMR and (b) ¹³C NMR spectra of 2b.

Because the ring strains of the oxirane-type monomers of **1a** and **1b** were larger than the tetrahydrofuran-type monomers of **3a** and **3b**, **2a** and **2b** were hyperbranched polytetritols without the 2,3-anhydroerythritol and 2,3-anhydro-L-threitol units as the anhydrotetritol units,¹¹ though the hyperbranched polytetritol prepared from **3a** and **3b** contained 1,4-anhydroerythritol and 1,4-anhydro-L-threitol as the anhydrotetritol units.³

Monomers **1a** and **1b** have an epoxy and two hydroxy groups in a molecule, so that they are classified as latent AB₃-type monomers. The degrees of branching (DBs) prepared from the AB₃-type monomers were determined from the ratio of the terminal unit using Frey's equation, as shown in the Experimental Section. ¹⁰ The ratio of the terminal unit in **2a**, $T_a/(D_a + sD_a + L_a + T_a)$, which was estimated from the ¹³C NMR measurement, was 31.0%. Thus, the DB of **2a** was determined to be 0.47, as summarized in Table 1. For polymer **2b**, the ratio of the terminal unit, $T_b/(D_b + sD_b + L_b + T_b)$, was 30.0%, and the DB was 0.45. Although the DBs of hyperbranched carbohydrate polymer prepared from **3a** and **3b** varied in the range of 0.22–0.46, the DBs did not change according to the polymerization condition.³

Three-Dimensional Property. In general, highly branched polymers, such as dendrimers, star polymers, and hyperbranched polymer, are known to have spherical conformations in solution. 12a,b,13 Thus, the $M_{\rm w,SLS}$ values were smaller than the weight-average molecular

2,3-*L*_a

weight $(M_{\text{w.SEC}})$ values estimated using size exclusion chromatography (SEC); i.e., the $M_{\rm w,SLS}$ values (2.67 \times $10^5 - 3.20 \times 10^6$) were ca. $2.57 \times 10^2 - 1.44 \times 10^3$ times larger than the $M_{\rm w,SEC}$ values (1.04 \times 10³–2.77 \times 10³). On the contrary, for the linear polymer $(2\rightarrow 3)$ -DLthreitol, the $M_{\rm w,SLS}$ value of 1080 was somewhat larger than the $M_{\rm w,SEC}$ value of 890. For the solution viscosity, a linear dependence between the reduced viscosity and the polymer concentration was observed, as shown in Figure 4, indicating that no aggregation occurred for 2a and **2b** in an aqueous NaNO₃ solution (0.2 mol L^{-1}). Therefore, the measured $M_{\rm w,SLS}$ values should correspond to the absolute molecular weights. The solution viscosity of 2a and 2b was extremely low, and the intrinsic viscosity ($[\eta]$) was in the range of 0.019-0.025 $dL g^{-1}$, as listed in Table 1.

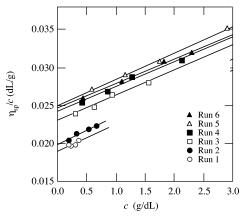


Figure 4. Viscosity vs concentration plots for 2a and 2b.

The relationship between $[\eta]$ and the molecular weight (M) can be described by the Mark-Houwink equation of $[\eta] = KM^a$. Figure 5 shows the doublelogarithmic plots of $[\eta]-M_{\text{w,SLS}}$. The value of the slope, which is *a* in the Mark–Houwink equation, was 0.105. This small value was consistent with a highly branched compact structure. 13 Therefore, these results of the SEC, SLS, and solution viscosity measurements suggested that 2a and 2b were highly branched spherical molecules, i.e., hyperbranched carbohydrate polymers.

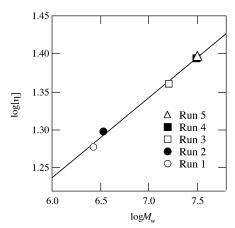


Figure 5. Logarithmic plots of $[\eta]-M_{\text{w,SLS}}$ for **2a**.

Polymerization Mechanism. Hyperbranched carbohydrate polymers 2a and 2b mainly consisted of DLthreitol and erythritol repeating units, respectively, as shown in Chart 2 and Chart 3, respectively. The polymerization, therefore, proceeded through a mechanism similar to the ring-opening multibranching po-

Scheme 3. A Proposed Mechanism for Ring-Opening Multibranching Polymerization with Proton-Transfer Reaction

Initiation reaction

Proton transfer reaction

lymerization of glycidol, which was reported by Kubisa and Penczek.^{1,2} Scheme 3 represents the proposed mechanism for the polymerization of 1a. In the polymerization system, the oxygen atom of the epoxy group of 1a was protonated, and the protonated 1a reacted with the oxygen atom of the epoxy group from another monomer, thus leading to 2,3- L_a . The branched unit for 2a should be produced by the proton-transfer reaction of the hydroxy groups in the polymer chain with the oxonium cation end of the other polymer chain. The polymerization of 1b proceeded through a mechanism similar to that for 1a to produce the hyperbranched carbohydrate polymer, 2b.

Conclusions

The polymerization of 2,3-anhydroerythritol (1a) and 2,3-anhydro-DL-threitol (1b) proceeded through the ringopening reaction with a proton-transfer reaction mechanism, i.e., the ring-opening multibranching polymerization. The resulting polytetritols free from cyclic units were the hyperbranched carbohydrate polymers 2a and 2b essentially consisting of DL-threitol and erythritol units, respectively, which differed from the hyperbranched polytetritol prepared from 1,4-anhydroerythritol (3a) and 1,4-anhydro-L-threitol (3b). The weightaverage molecular weight values ($M_{w,SLSS}$) were remarkably higher than those of the hyperbranched carbohydrate polymer prepared by the ring-opening multibranching polymerization of 3a and 3b. The threedimensional properties characterized by size exclusion chromatography, static light scattering, and the viscosity measurements indicated that 2a and 2b should be spherical molecules.

Acknowledgment. The authors thank the Akiyama Foundation (Japan) and the Hayashi Memorial Foundation for Female Natural Scientists for their financial support.

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